

# The Influence of Newly Synthesised Fenpropimorph Derivatives on Some Pathogen Yeasts

Emília Breierová<sup>a,\*</sup>, Ján Šajbidor<sup>b</sup>, and Martin Lamačka<sup>b</sup>

<sup>a</sup> Culture Collection of Yeasts, Institute of Chemistry, Slovak Academy of Sciences, Dúbravská cesta 9, 842 38 Bratislava, Slovakia. Fax: +421-7-59410222. E-mail: chememi@savba.sk

<sup>b</sup> Department of Biochemical Technology, Slovak University of Technology Radlinského 9, 812 37 Bratislava, Slovakia

\* Author for correspondence and reprint requests

Z. Naturforsch. **56c**, 53–57 (2001); received July 28/September 12, 2000

Fenpropimorph Derivatives, Ergosterol, Lipids

The effect of minimum inhibitory concentrations (MICs) of six novel fenpropimorph derivatives on lipid and sterol composition of *Candida albicans*, *Cryptococcus neoformans*, *Malassezia pachydermatis* and *Malassezia furfur* was investigated. The MICs for the most effective derivatives were found in the range from 3.7 to 56.7  $\mu\text{M}$  and were 2–3 times lower compared to the commercial fungicide bifonazol. The more efficient fenpropimorph derivatives were the piperidine derivative for *C. albicans* and the allylamine derivative for *Cr. neoformans*, *M. pachydermatis* and *M. furfur*. The inhibitor in the growth medium reduced the unsaturation index of the total lipid content in *M. furfur* and *C. albicans*.

## Introduction

The increasing incidence of yeast infections has greatly stimulated the interest in the development of new antimycotic drugs affecting the sterol biosynthesis pathway. The most frequently used fungicides are azoles and morpholines. The target enzyme for azoles is lanosterol demethylase (Joseph-Horne and Hollomon, 1997), morpholines and piperidines inhibit the  $\Delta^{8-7}$  isomerase and  $\Delta^{14}$ -reductase (Marcireau *et al.*, 1992). Although attack of ergosterol biosynthesis is usually the primary reason of growth inhibition, unsaturated sterols can replace ergosterol in the membranes. So, a fungistatic effect can not only be due to accumulation of abnormal sterols in treated cells, but is linked with other adverse influences (Marcireau *et al.*, 1990). In accordance with this suggestion Georgopapadakou *et al.* (1987) showed that changes in membrane fatty acids induced by fungicides rather than decreased ergosterol were responsible for growth inhibition of *Candida albicans*. All yeasts used belong to the most important opportunistic pathogens for immunocompromised humans or animal hosts (Groshek, 1998) *C. albicans* is the main cause of superficial and systematic mycoses, and *Cryptococcus neoformans* is an important pathogen for individuals with AIDS and other immu-

nocompromising diseases (Kelly *et al.*, 1999; Bujdaková *et al.*, 1999). The lipophilic yeast *Malassezia pachydermatis* and lipid-dependent species *Malassezia furfur* are part of the normal cutaneous microflora of most warm-blooded vertebrates (Boerhout *et al.*, 1998; Guillot and Bond, 1999).

The objective of the present study was to compare the antifungal activity of six fenpropimorph derivatives including allylamines, piperazines and piperidines. Their inhibitory effect was compared with the azol fungicide bifonazol.

## Materials and Methods

The following pathogen yeasts were used: *Candida albicans* CCY 29–3-101, CCY 29–3-102, *Cryptococcus neoformans* CCY 17–1-2, *Malassezia furfur* CCY 85–2-1 and two strains of *Malassezia pachydermatis* (CCY 85–1-5, CCY 85–1-10). The strains were obtained from the Culture Collection of Yeasts (Institute of Chemistry, SAS, Bratislava, Slovakia).

The cultures were grown on malt agar at 37 °C, only *M. furfur* was cultivated on Dixon's modified medium (7 g malt extract, 1 ml Tween 40, 0.25 ml glycerol, 1 g peptone, 0.01 g yeast extract, 0.05 g stearic acid and 2 g agar in 100 ml medium). Inhi-

0939–5075/2001/0100–0053 \$ 06.00 © 2001 Verlag der Zeitschrift für Naturforschung, Tübingen · www.znaturforsch.com · D



Dieses Werk wurde im Jahr 2013 vom Verlag Zeitschrift für Naturforschung in Zusammenarbeit mit der Max-Planck-Gesellschaft zur Förderung der Wissenschaften e.V. digitalisiert und unter folgender Lizenz veröffentlicht: Creative Commons Namensnennung-Keine Bearbeitung 3.0 Deutschland Lizenz.

Zum 01.01.2015 ist eine Anpassung der Lizenzbedingungen (Entfall der Creative Commons Lizenzbedingung „Keine Bearbeitung“) beabsichtigt, um eine Nachnutzung auch im Rahmen zukünftiger wissenschaftlicher Nutzungsformen zu ermöglichen.

This work has been digitalized and published in 2013 by Verlag Zeitschrift für Naturforschung in cooperation with the Max Planck Society for the Advancement of Science under a Creative Commons Attribution-NoDerivs 3.0 Germany License.

On 01.01.2015 it is planned to change the License Conditions (the removal of the Creative Commons License condition “no derivative works”). This is to allow reuse in the area of future scientific usage.

bition effects of the newly synthesised fungicides (Fig. 1) prepared according to Veverka *et al.*, 1990, was compared with bifonazol (Bayer AG, Germany). Paper discs (0.5 cm in diameter) were impregnated with 10 µl of inhibitor solution in 96% ethanol at the final amounts of 250, 125 and 50 µg per disc. The discs were placed on agar surface covered with culture suspension. The yeast strains were incubated at 37 °C for 48–72 h. Zones of growth inhibition (mm in diameter) were measured for each amount of fungicide. For the most effective inhibitors the minimum inhibitory concentrations (MICs) were determined by a microdilution method according Carillo-Munoz and Tur-Tur (1997). All determinations were made in triplicate. Cells used for sterol and fatty acid analysis were grown on malt agar with a suspended fungicide concentration of 5 µg·ml<sup>-1</sup> in the medium. Lipids were extracted from lyophilised yeasts with a chloroform / methanol mixture (2:1 v/v) (Folch *et al.*, 1957). Fatty acids were liberated by alkaline hydrolysis, methylated with diazomethane (Schlenk and Gellerman, 1960) and analysed according to Bohov *et al.* (1997). Sterols were determined by GC (Pesti *et al.*, 1997) and HPLC (Lamačka and Šajbidor, 1997; Steel *et al.*, 1989).

## Results and Discussion

The tested fenpropimorph derivatives (Fig. 1) showed differences between basidiomycete species *M. pachydermatis*, *M. furfur*, and *Cr. neoformans* and the ascomycete *C. albicans* (Table I). Allylamine R<sub>1</sub> and piperazine R<sub>2</sub> derivatives are weak inhibitors for all tested strains. The piperidine derivative R<sub>3</sub> was effective against *C. albicans* and *Malassezia* sp. Similarly, R<sub>4</sub> was inhibitory for two isolates of *M. pachydermatis*, but an effect on *M. furfur* was not detected. R<sub>5</sub> and R<sub>6</sub> strongly influenced the growth of *Malassezia* and *Cryptococcus*, but not *C. albicans*. Comparison of the MIC showed, that the allylamine R<sub>5</sub> derivative was the effective fungicide for *M. pachydermatis*, *M. furfur* and *Cr. neoformans*. Similarly, R<sub>3</sub> was more potent than the reference for both *Candida* strains (Table II).

As the target of all tested fungicides are enzymes of the sterol pathway, changes in the content of ergosterol were investigated. It is evident that the presence of inhibitor caused its decrease in all samples, but a direct correlation between the extent of ergosterol reduction and MIC was not observed. The decline in ergosterol, di-

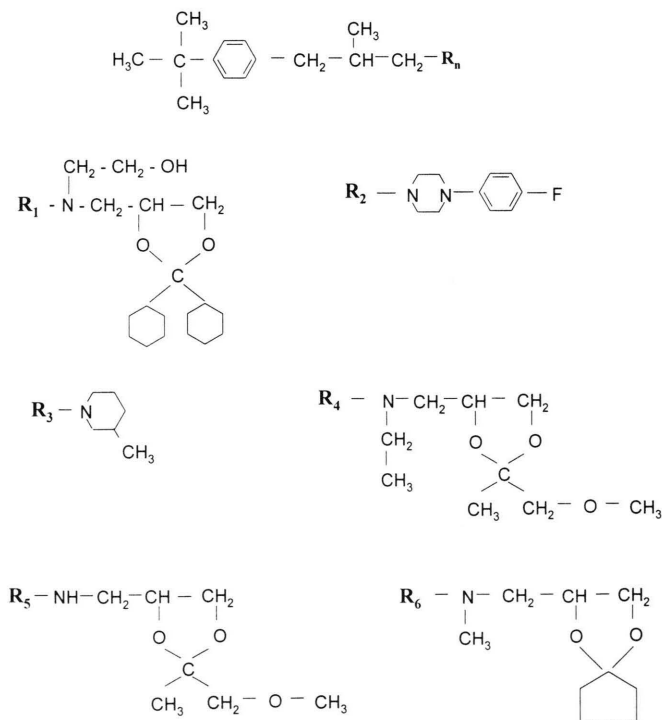


Fig. 1. Chemical structures of newly synthesised inhibitors.

Table I. Influence of three concentrations of inhibitors on growth of some pathogenic yeasts.

Inhibitor	c µg/disc	Yeast strain CCY					
		<i>M. pachy- dermatis</i> 85-1-5	<i>M. pachy- dermatis</i> 85-1-10	<i>M. furfur</i> 85-2-1	<i>Cr. neo- formans</i> 17-1-2	<i>C.</i> albicans 29-3-101	<i>C.</i> albicans 29-3-102
		Inhibition <sup>a</sup>					
<b>R<sub>1</sub></b> (allylamine)	50	—	—	—	—	—	—
	125	+	—	—	—	—	—
	250	+	—	—	—	—	—
<b>R<sub>2</sub></b> (piperazine)	50	—	—	—	—	—	—
	125	+	—	—	—	—	—
	250	+	—	—	—	—	—
<b>R<sub>3</sub></b> (piperidine)	50	—	—	—	—	—	+
	125	++	++	—	—	+	+
	250	++	++	—	—	++	++
<b>R<sub>4</sub></b> (allylamine)	50	—	—	—	—	—	+
	125	++	++	—	—	—	+
	250	++	++	—	—	+	+
<b>R<sub>5</sub></b> (allylamine)	50	++	++	++	++	—	—
	125	+++	+++	++	++	—	—
	250	+++	+++	+++	++	—	+
<b>R<sub>6</sub></b> (allylamine)	50	++	++	+	+	—	—
	125	++	++	++	+	—	—
	250	++	++	++	+	—	+
bifonazol	50	++	++	+	+	—	—
	125	++	++	++	+	—	—
	250	++	++	++	++	—	+

c – concentration of inhibitor per disc; <sup>a</sup> – zone of inhibition (mm in diameter): 0 (–), 0–10 (+), 10–20 (++), 20–30 (+++).

Table II. Ergosterol content of yeast cultivated without (–) and with (+) inhibitors at the minimal inhibition concentration (MIC) of the most effective inhibitors and bifonazol.

Yeast strain CCY	MIC (µM)		Ergosterol content (µg · mg <sup>-1</sup> DC)	
	Bifonazol	inhibitor <sup>a</sup>	–	+
<i>M. pachydermatis</i> 85-1-5	26.5	9.2	1.5	0.3
<i>M. pachydermatis</i> 85-1-10	28.7	7.7	1.2	0.5
<i>M. furfur</i> 85-2-1	8.6	3.7	0.5	0.03
<i>Cr. neoformans</i> 17-1-2	57.1	17.5	0.4	0.3
<i>C. albicans</i> 29-3-101	115.3	56.7	2.2	0.09
<i>C. albicans</i> 29-3-102	98.9	43.1	2.2	0.4

<sup>a</sup> Inhibitor R<sub>5</sub> was used for 85-1-5, 85-1-10, 85-2-1 and 17-1-2, inhibitor R<sub>3</sub> was tested against 29-3-101 and 29-3-102 strains; DC – dry cell weight; (–) absence of inhibitors; (+) presence of inhibitors.

hydroergosterol and episterol in *C. albicans* was accompanied with the increasing of zymosterol, ignosterol, and fecosterol (Table III). Accumulation of zymosterol could be related to the reduction of C24 sterol methyl transferase activity (Barrett-Bee and Dixon, 1995), probably the high

Table III. The influence fenpropimorph derivative R<sub>3</sub> on the profile of sterols using the more resistant strain *Candida albicans* CCY 29-3-102.

Sterol	Sterol content (µg · g <sup>-1</sup> DC)	
	–	+
Ergosterol	802	162
Zymosterol	10	258
Dihydroergosterol	93	2
Ignosterol	6	478
Lanosterol	12	38
Fecosterol	4	59
Episterol	75	3

DC – dry cell weight; (–) absence of inhibitors, (+) presence of inhibitors.

Table IV. Fatty acid composition of *M. furfur* and *C. albicans* cultivated without (–) and with (+) the most effective inhibitors ( $R_5$  for *M. furfur* and  $R_3$  for *C. albicans*).

Fatty acids	<i>Malassezia furfur</i> CCY 85–2-1		<i>Candida albicans</i> CCY 29–3-102	
	–	+	–	+
Fatty acid content % DC				
12:0	0.5	9.6	0.1	0.1
13:0	ND	0.5	ND	ND
14:0	2.8	11.2	0.7	0.7
15:0 ai	8.6	ND	ND	ND
14:1 n-5	ND	0.4	0.1	ND
15:0	0.8	2.4	0.2	0.3
16:0 i	2.6	ND	ND	ND
16:0	44.0	54.6	16.9	12.0
16:1 n-7t	0.3	0.5	0.1	0.2
16:1 n-9c	1.7	0.2	0.2	0.3
16:1 n-7c	1.9	3.7	5.7	9.9
17:0 ai	2.2	0.2	0.6	0.2
17:0	0.9	2.0	1.1	0.7
17:1 n-8c	0.3	0.2	1.4	1.8
18:0	15.9	5.8	7.0	6.3
18:1 n-9t	0.4	ND	0.2	0.3
18:1 n-7t	ND	0.2	0.1	0.2
18:1 n-9 c	10.4	0.6	43.6	41.8
18:1 n-7c	0.6	2.9	0.7	0.8
18:2 n-6c	4.3	3.7	15.3	19.0
20:0	1.1	0.1	0.3	0.3
18:3 n-3c	ND	ND	1.7	2.3
20:2 n-6c	0.2	1.3	0.1	0.1
22:0	0.1	ND	0.2	0.2
24:0	0.2	ND	2.7	1.8
25:0	ND	ND	0.3	0.4
26:0	0.2	ND	0.6	0.4
<b>I.U</b>	<b>0.2464</b>	<b>0.1866</b>	<b>0.8812</b>	<b>1.0024</b>

DC – dry cell weight, ai – ante-iso, i – iso, I.U – index of unsaturation calculated according Kates and Baxter, 1962, ND – not determined.

content of ergosterol was the consequence of  $\Delta^{14}$ -reductase inhibition (Marcireau *et al.*, 1992). Increase of fecosterol was always accompanied with  $\Delta^{8-7}$  isomerase inhibition (Kelly *et al.*, 1994).

Cultivation in the presence of fungicide induced adaptive changes not only in the sterol profile, but also in fatty acid content and composition (Table IV). It is interesting that fungicides caused only a slight decrease of the total fatty acid level amount in both tested strains. Although growth inhibition is usually accompanied with lipid reduction (Barrett-Bee and Dixon, 1995), the increase of the neutral lipid level can also be a consequence of ergosterol disappearance (Hitchcock *et al.*, 1987). GC analyses showed that the content of individual fatty acids in *Malassezia furfur* and *Candida albicans* were quite different. The main fatty acid in *M. furfur* was palmitic in contrary with *C. albicans*, in which oleic acid was detected as the most abundant structure. *M. furfur* shows the inhibitory effect by stearic and oleic acid reduction with concomitant increase of palmitic acid. On the other hand, cultivation of *C. albicans* in the presence of the  $R_3$  derivative decreased both, palmitic and oleic acid as well. Decline of these was compensated by accumulation of palmitoleic, linoleic and linolenic acid. Generally, the adaptation response of *M. furfur* included a slight reduction of lipid unsaturation unlike *C. albicans* where the opposite trend was observed.

The results demonstrate that fenpropimorph derivatives exhibit potent mycostatic and mycotoxic activity against *Cr. neoformans*, *Malassezia pachydermatis* and *M. furfur* with the  $R_5$  - allylamine derivative and against *C. albicans* with the  $R_3$  - piperidine derivative.

#### Acknowledgements

We would like to thank Dr. M. Veverka for supplying tested derivatives. This work was supported by grant No. 1/6252 and 2/4145 from the Slovak Grant Agency.

- Barrett-Bee K. and Dixon G. (1995), Ergosterol biosynthesis inhibition: a target for antifungal agents. *Acta Biochim. Pol.* **42**, 465–479.
- Boerhout T., Kamp M. and Gueho E. (1998), Molecular typing of *Malassezia* species with PFGE and RAPD. *Medical Mycology* **36**, 365–372.
- Bohov P., Sebková E., Gasperikova D., Langer P. and Klimes I. (1997), Fatty acid composition in fractions of structural and storage lipids in liver and skeletal muscle of hereditary hypertriglyceridemic rats. *Ann. N. Y. Acad. Sci.* **827**, 494–509.
- Bujdaková H., Lell P. C., Gruber A., Langgartner M., Spötl L., Kurišová S., Klobušický M., Dierich M. P. and Würzner R. (1999), The influence of subinhibitory concentrations of conventional and experimental antifungal drugs on the expression of the iC3b binding protein in *Candida albicans* strains during filamentation. *FEMS Immun. Med. Microbiol.* **26**, 1–10.
- Carillo-Munoz A.-J. and Tur-Tur C. (1997), Comparative study of antifungal activity of sertaconazole, terbinafine, and bifonazole against clinical isolates of *Candida* spp., *Cryptococcus neoformans* and dermatophytes. *Chemotherapy* **43**, 387–392.
- Folch J., Lees M. and Sloane-Stanley G.-H. (1957), Simple method for the isolation and purification of total Lipids from animal tissues. *J. Biol. Chem.* **226**, 497–509.
- Georgopapadakou N.-H., Dix B.-A., Smith S.-A., Freudenberg J. and Funke P.-T. (1987), Effect of antifungal agents on lipid biosynthesis and membrane integrity in *Candida albicans*. *Antimicrob. Agents Chemother.* **31**, 46–51.
- Groshek P.-M. (1998), *Malassezia pachydermatis* infections. *N. Engl. J. Med.* **339**, 270–271.
- Guillot J. and Bond R. (1999), *Malassezia pachydermatis*: a review. *Medical Mycology* **37**, 195–306.
- Hitchcock C.-A., Barrell-Bee K.-J. and Russell N.-J. (1987), The lipid composition and permeability to azole of an azole- and polyene-resistant mutant of *Candida albicans*. *J. med. Vet. Mycol.* **25**, 29–37.
- Joseph-Horne T. and Hollomon D.-W. (1997), Molecular mechanisms of azole resistance in fungi. *FEMS Microbiol. Lett.* **149**, 141–149.
- Kates M. and Baxter R.-M. (1962), Lipid composition of mesophilic and psychophilic yeasts (*Candida* species) as influenced by environmental temperature. *Can. J. Biochem. Physiol.* **40**, 1213–1227.
- Kelly S.-L., Lamb D.-C. and Kelly D.-E. (1999), Y132H substitution in *Candida albicans* sterol 14 $\alpha$ -demethylase confers fluconazole resistance by preventing binding to haem. *FEMS Microbiol. Lett.* **180**, 171–175.
- Kelly D.-E., Rose M.-E. and Kelly S.-L. (1994), Investigation of the role of sterol  $\Delta^{8-7}$  isomerase in the sensitivity of *Saccharomyces cerevisiae* to fenpropimorph. *FEMS Microbiol. Lett.* **122**, 223–226.
- Lamačka M. and Šajbidor J. (1997), Ergosterol determination in *Saccharomyces cerevisiae*. Comparison of different methods. *Biotechnol. Techniques* **11**, 723–725.
- Marcireau C., Guilloton M. and Karst F. (1990), *In vivo* effects of fenpropimorph on the yeast *Saccharomyces cerevisiae* and determination of the molecular basis of the antifungal property. *Antimicrob. Agents Chemother.* **34**, 989–993.
- Marcireau C., Guyonnet D. and Karst F. (1992), Construction and growth properties of a yeast strain defective in sterol 14-reductase. *Curr. Genet.* **22**, 267–272.
- Pesti M., Horváth L., Vígh L. and Farkas T. (1985), Lipid content and ESR determination of plasma membrane order parameter in *Candida albicans* sterol mutants. *Acta Microbiol. Hung.* **32**, 305–313.
- Steel C.-C., Baloch R.-I., Mercer E.-I. and Baldwin B.-C. (1989), The intracellular location and physiological effects of abnormal sterols in fungi grown in the presence of morpholine and functionally related fungicides. *Pestic Biochem Physiol* **33**, 101–111.
- Veverka M., Štibrányi L. and Szemes F. (1990), The method for the preparation of phenylpropylamine derivatives. *CS. Pat. Appl.* 11790.